

**RNA and DNA Consist of Polymerized Nucleotides**

5'-end

3'-OH

2'-OH

3'-end

Ribonucleic acid (RNA)

Deoxyribonucleic acid (DNA)

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**Components of RNA and DNA**

AMP

dAMP

CMP

dCMP

GMP

dGMP

UMP

dTMP

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**RNA Is More Labile Than DNA**

RNA is labile in a water solution because the phosphate groups can form phosphodiester bonds with both the 2' and 3'-OH groups of ribose.

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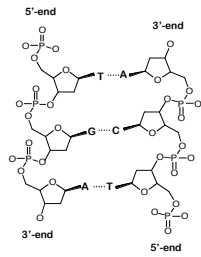
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### The Strands of DNA are Anti-Parallel



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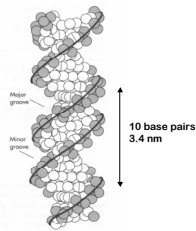
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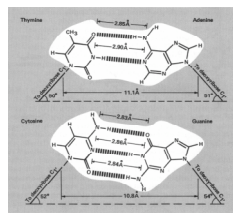
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### Structure of Double-Stranded DNA

B-form of DNA



Base-pairing of B DNA



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### The Size of Cellular DNA

*Escherichia coli* (a bacterium) has one DNA molecule (chromosome) with a size of ca.  $10^9$  nucleotides

*Homo sapiens* cells have 46 chromosomes, each consisting of one DNA molecule, with a total size of ca.  $10^9$  nucleotides

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**Nuclear DNA Forms Chromatin**

Uppsala University

Histones are among the most conserved proteins in eukaryotes

Chromatin fiber

Linker histones

Long range fiber-fiber interactions

30nm fiber

Short range internucleosomal interactions

G1 chromatid

Beads-on-a-string

Nucleosome

DNA

Core histone tail domain

The nucleosome core is composed of an octamer of histone molecules. Two each of the histones H2A, H2B, H3 and H4. H1 is a linker histone.

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**Synthesis of DNA**

Uppsala University

Enzymes catalysing the synthesis of DNA: DNA polymerases.

DNA polymerases have some general properties

- DNA molecules are extended at the 3'-end. DNA is synthesised in the 5' to 3' direction
- Deoxynucleoside triphosphates (dATP, dCTP, dGTP, dTTP) are used as building blocks
- The polymerisation reaction is directed by A-T, G-C base pairing to a template strand

Primer

5'

3'

Template strand

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**Polymerisation of deoxynucleotides**

Uppsala University

Growing strand

Primer

Active site of DNA polymerase

Mg<sup>2+</sup>

Template strand

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### Fidelity of DNA synthesis

In mammals, the error frequency of DNA synthesis must be  $<10^{-9}$

The base pairing mechanism leads to one error in  $\sim 10^4$  polymerised nucleotides

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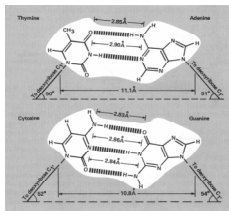
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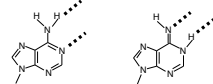
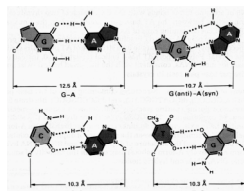
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### Reasons for Incorrect Base-pairing



Correct base-pairing



Tautomer of A can base pair with C instead of T

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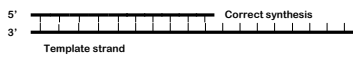
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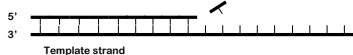


### Correction of Replication Errors

#### Proofreading activity of DNA polymerases



3' to 5' exonuclease activity of DNA polymerase catalyses the removal by hydrolysis of 3'-terminal nucleotides that are not base-paired to the template strand.



With this mechanism the error frequency of DNA synthesis is decreased to  $\sim 10^{-6}$ .

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**Correction of Replication Errors**

DNA polymerase I from *E. coli* has two catalytic activities, although the enzyme consists of a single polypeptide chain.

The diagram shows DNA polymerase I (Pol I) bound to a primer-template junction. On the left, the enzyme is in its synthesis mode, with the 3'OH of the primer being extended. On the right, it is in its proofreading mode, with the 3'OH of the primer being removed. The enzyme is depicted with hand-like regions: Fingers, Thumb, Palm, and Exo (exonuclease active site).

View of a DNA polymerase bound to a primer-template junction in its synthesis (left) and proofreading (right) modes. Pol denotes the polymerase active site; Exo, the 3' to 5' exonuclease active site.

From Baker & Bell (1998) Cell, 92:295

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**Correction of Replication Errors**

Postreplication mismatch repair

The diagram shows a DNA double strand with a methylated base (marked with an asterisk) on the template strand. A mismatch is shown between the template and the newly synthesized strand. The mismatch repair system recognizes the mismatch and removes the nucleotide in the unmethylated (newly synthesized) strand.

In DNA, some of the C (and A) bases are methylated. Methylation is a slow process. Newly replicated DNA is undermethylated.

Mismatch repair enzymes recognise mismatched nucleotides and remove the nucleotide in the undermethylated strand.

The mismatch repair system increases the fidelity of DNA replication to  $\sim 10^{-9}$ .

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**Replication of Chromosomal DNA**

Semiconservative DNA replication

The diagram shows a single DNA molecule (two strands) replicating to form two DNA molecules, each with one original strand and one newly synthesized strand.

A half-replicated DNA molecule

What happens at the replication fork?

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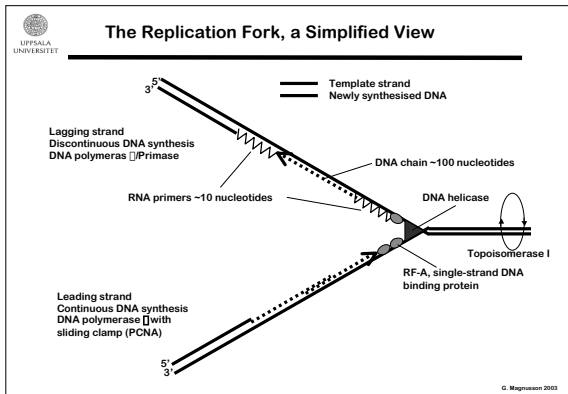
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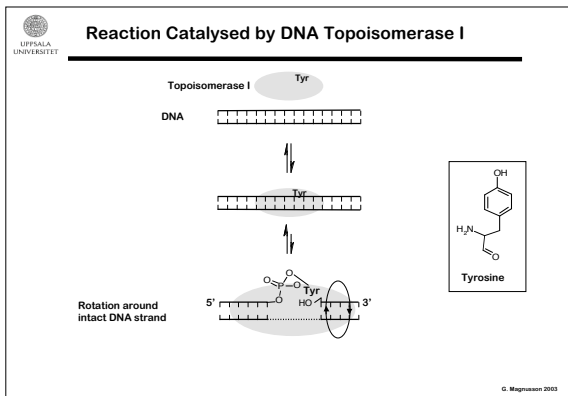
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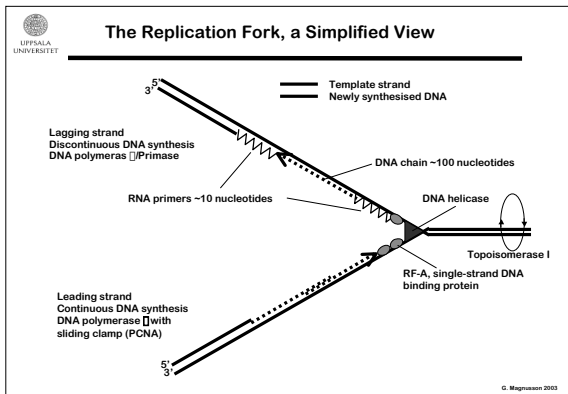
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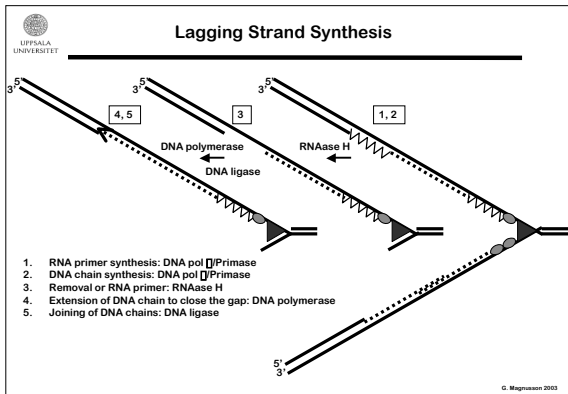
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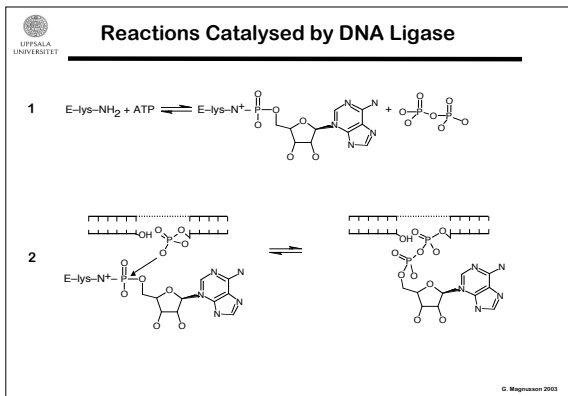
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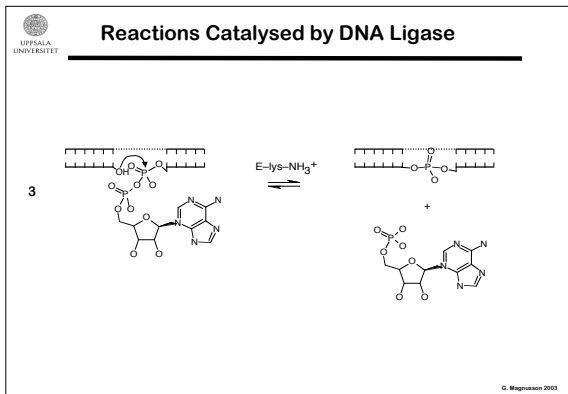
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**Replication of Chromosomal Ends (Telomers)**

Loss of genetic information at chromosomal ends is prevented by the addition of nontemplated nucleotides by telomerase

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**Mechanism of Telomerase Activity**

Telomerase is a DNA polymerase that uses an RNA template. This template is a subunit of the enzyme and consists of an RNA molecule with a AAUCCC sequence repeat.

In the new-born, the telomeres have a length  $\sim 10^4$  nucleotides with ca.  $10^2$  unpaired nucleotides at the 3'-end.

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**Initiation of DNA-Synthesis at an Origin**

From Bogdan et al. (2000) J. Cell. Physiol. Vol. 184, 139

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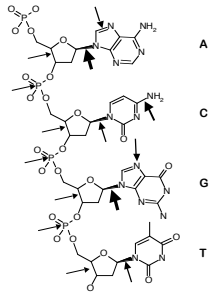


### Frequent Spontaneous DNA Damage

Frequency of individual lesions of nucleotides is suggested by the size of the arrows.

Depurination is the most frequent lesion  $\sim 10^4$ /day/cell.

Deamination of C to form U occurs  $\sim 10^3$  times per day in a cell.



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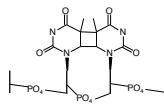
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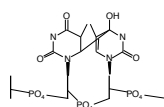
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### UV-Induced Damage to DNA

Absorption of UV-radiation (200–320 nm) by DNA induces two major types of potentially mutagenic lesions. Skin cells not protected by pigmentation accumulate  $\sim 10^4$  lesions per hour when exposed to strong sun-light.



Cyclobutane thymine dimer kinks DNA  $7^\circ-9^\circ$



Thymine (6-4) pyrimidone photoproduct kinks DNA  $40^\circ$

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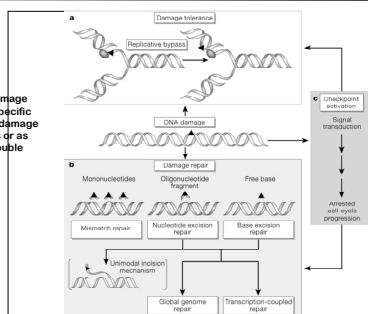
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### Mechanisms of DNA Repair

In the cell, DNA damage is recognised by specific proteins either as damage to individual bases or as distortion of the double helix.



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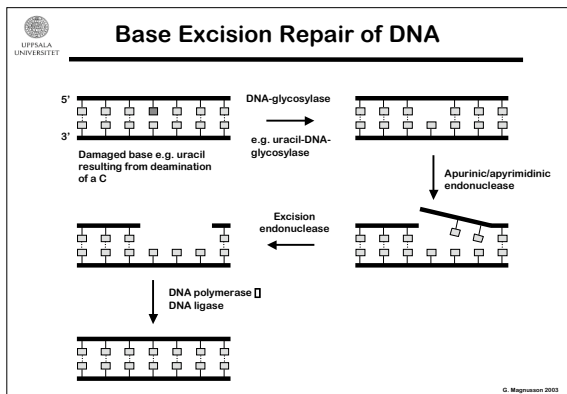
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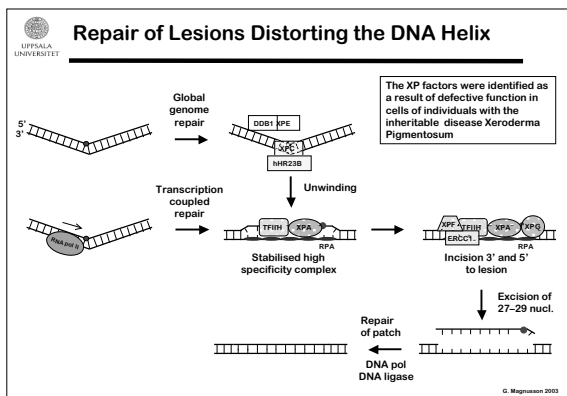
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- Ames' Mutagen Test**
- Salmonella bacteria requiring histidine for growth
  - Use strains with several different types of mutations in the *his* genes
  - Treat bacteria with test chemical, directly or after metabolic activation in liver cell extract
  - Plate bacteria on agar in histidine deficient medium
  - Count colonies and calculate mutation frequency
- © Magnusson 2003

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## Mouse Carcinogen Test

- Treat animals with test substance. For substances with low potency, use high doses or many animals
- $TD_{50}$  (mg/kg bodyweight/day during lifetime) calculated as the dose of chemical that would decrease the number of tumour-free animals by 50%
- Tumours have to be analysed by a pathologist
- $TD_{50}$  for various substances has a range of  $>10^7$

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## Risk Assessment

- All environmental risks cannot be avoided
- There is no inherent difference in the toxicity of natural and synthetic substances
- Human exposure has to be considered in the assessment of risk
- HERP (human exposure rodent potency). Percentage of the rodent potency ( $TD_{50}$ ) received by a human during lifetime exposure

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## HERP

### HERP (%)<sup>a</sup>

- **3.6** Beer, 500 ml (ethyl alcohol, 23 ml/day)
- **0.4** Typical indoor air (formaldehyde, 600  $\mu$ g/day)
- **$1 \times 10^{-6}$**  Lindane<sup>b</sup> (32 ng/day)

<sup>a</sup>A HERP value of 100 means that the exposure is identical to the  $TD_{50}$  in rodent tests.

<sup>b</sup>Lindane is a mutagenic insecticide that is banned from use in Swedish agriculture.

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